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Author's Affiliation:

1. Department of Clinical Laboratory sciences, Faculty of Applied Medical Sciences, Umm Al-Qura University, Makkah - Kingdom of Saudi Arabia

2. Department of Medical Laboratory Sciences, Mohammed Al-Mana College of Medical Sciences, Al Safa, Dammam - Saudi Arabia

Corresponding Author:

Afnan Mohammed Shakoori Email: amshakoori@uqu.edu.sa

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and GPS 6.0

You are viewing the latest version of this article having minor corrections related to the use of English language.

Structural interpretation of the NS3 helicase ATP binding domain of Zika virus (ZIKV)

Afnan Mohammed Shakoori^{1*}, Nida Alsaffar²

Abstract

ackground: The Zika virus, a mosquito-borne virus, was discovered in Uganda and quickly spread to Asia and the Pacific. Zika is mainly transmitted by the bite of an infected Aedes species mosquito (Aedes aegypti and Aedes albopictus). These mosquitos bite both during the day and at night.

Methods: Numerous tools (SMART, Pfam, InterProScan and Scan Prosite, BepiPred-2.0 server, I-TASSER, PROCHECK, phyre 2, protparam, and GPS 6.0) were used to elaborate the content in this publication.

Results: The results show that HABD (187 amino acids) is a DEAH-Box RNA helicase. We also used the Ramachandran plot to validate the epitope peptides and structure modeling. We also discovered the amino acid composition and various residues/parentages in the HABD protein phosphorylation site prediction and PK-specific phosphorylation sites (p-sites).

Conclusion: The helicase ATP binding domain (HABD), HABD protein ATP binding sites, and epitope binding peptides are discussed in this work. There are five atoms in total: nitrogen, sulfur, hydrogen, carbon, and oxygen. Hydrogen atoms (1476) provide the most to the composition. We created a graph representing the protein's predicted phosphorylation sites (p-sites). Aside from the standard statistics, GPS 6.0 may identify PK-specific p-sites hierarchically. GPS 6.0 could be a valuable service for further phosphorylation study.



Introduction

Zika virus (ZIKV) is a member of the Flaviviridae family and the Flavivirus genus. With 53 species, the flavivirus genus is the largest in the Flaviviridae family and can be further subdivided into non-vector, tick-borne, and mosquito-borne clusters [1,2]. Mosquito-borne flaviviruses such as ZIKV, dengue virus (DENV), West Nile virus (WNV), yellow fever virus (YFV), and Japanese encephalitis virus (JEV) have been a growing public health concern in the recent decade due to an increase in global incidence [1,2]. ZIKV traveled across the Pacific Ocean in May 2015 and was introduced in Brazil, where it caused over one million cases [3]. As of May 2019, the virus had spread swiftly to 84 nations, territories, or subnational areas, posing a global public health threat [4]. ZIKV's genome is a 10.8-kb singlestranded positive-sense RNA molecule with a 100nucleotide 5' untranslated region (UTR), a single 10 kb open reading frame (ORF), and around 420 nucleotides in the 3' UTR. The ORF encodes a significant polyprotein precursor of 3423 amino acids, which is then co- and post-translationally cleaved into three structural (C, prM, and E) and eight nonstructural (NS1, NS2A, NS2B, NS3, S4A, NS4B, & NS5) proteins [5]. The N-terminal protease and C-terminal helicase domains of NS3 are structurally distinct functional domains [6]. Serine protease, NTPase, and RNA helicase (ATPase and Unwinding activity) are all found in NS3.

Antigen-antibody interaction is critical in immunological processes and reactions within the immune system, & B-cell epitopes are antigen regions recognized and bound by B-cell-produced antibodies [7]. Additionally, in the presence of particular crossreacting antibodies, B-cell epitopes can stimulate the humoral immune response [8,9]. The method of quickly identifying immunological epitopes is based on computational prediction, which employs advanced computations and an increasing epitope information base. The phosphorylation potential of ZIKV proteins has yet to be determined. Understanding the phosphorylation potential of viral proteins is critical since it aids in viral replication and interaction with host receptors [10]. This study provides molecular insight for future research into the structural and functional characteristics of the HABD protein. Protein phosphorylation is required for several biological functions. Protein phosphorylation is one of the most researched posttranslational modifications (PTMs) that control physical processes [11]. Protein kinases catalyze the reversible covalent attachment of a phosphate group on particular serine, threonine, and tyrosine residues of proteins [12, 13, 14]. Many in silico prediction approaches, in addition to experimental screening, have been developed to discover PK-specific phosphorylation sites (p-sites) efficiently and

efficiently. From 2004 to 2017, we created various GPS algorithms for p-sites and other PTM site predictions, ranging from version 1.0 to 4.0 [15, 16, 17, 18]. 2020, our team will produce GPS 5.0, combining position weight determination and score matrix optimization modules. GPS 5.0 could predict the p-sites of 479 human PKs inside the classical module [19]. Identifying potential phosphorylation sites and the protein kinases that bind to them is critical for understanding how many proteins work. This paper describes GPS 6.0, a prediction system that predicts phosphorylation sites in a kinase-family-specific manner. We can now explore phosphorylation-dependent processes and probe the kinases involved in controlling these functions since we have a predictor that effectively incorporates the context and sequence conditions that govern phosphorylation.

Methods

The arrangement of the helicase ATP binding domain (HABD)

The 187 amino acid sequence of the Zika Virus Helicase ATP binding domain protein detailed domain arrangement. One of the critical domains is the crucial (HABD) helicase ATP binding domain. For this study, the sequence of this HABD domain was retrieved from the NCBI genome database. We use SMART, Pfam, InterProScan, and Scan Prosite the most for manual domain categorization [20,21,22]. We also used the BepiPred-2.0 server to predict B-cell epitope peptides from the sequence [23].

I-TASSER is used to predict 3-D structure

The amino acids of the 187 Helicase ATP binding domain (HABD) sequence were obtained from a database in FASTA format. The 3-D (Threedimensional) model was created with the I-TASSER accessible service, which builds a three-dimensional model of the query sequence using multiple threading alignments and iterative structure assembly simulation [24]. The details for choosing this server were its accessibility, combined modeling technique, & success in the CASP competition. Threading, structural complex, model selection, refining, & structure-based efficient annotation are all general processes in the I-TASSER technique [25]. First, the evolutionary related protein was tested against the query sequence using PSI-BLAST, which generated a sequence profile that PSIPRED used to estimate secondary structure [26]. The request sequence was then LOMETS [27] routed by the illustrative PDB structure collection. Z-score was used to assess the excellence of template alignment, and the best one was chosen for further examination. The constant pieces in threading alignments were removed from an assembled structural model of aligned

regions in the following phase [24]. Unaligned region modeling accuracy is generally low, whereas threading aligned area modeling accuracy is high. Thus, these template fragments remain rigid throughout the simulation to get a high-resolution structure. For fragment assembly, the replica exchange Monte Carlo simulation technique was used [28]. C/side chain correlation, H-bonds, hydrophobicity, spatial restraints from threading templates [28], & sequence-based contact predictions from SVMSEQ [29] are all included in the refinement simulation-generated conformations [30]. The designated cluster centroids were employed over in the refinement phase to do additional fragment association simulation, which helps to reduce steric conflicts and refine the global topology of the cluster centroids. TM-align [31] was used to identify the PDB structures structurally near the cluster centroids. REMO [32] was used to create the final structural models, which employed cluster centroids from the second simulation round as input. In the final phase, the function of the three-dimensional model of the request protein was anticipated by comparing it to proteins with known structure & function in the PDB. The functional analogs were ordered according to their TM score, RMSD, sequence identity, and structural alignment coverage. The C-score (confidence score) measured the anticipated model's quality, which varied from 5 to 2. It is determined by threading alignment quality & the convergence of structural association refinement simulations.

Validation of structure models

The Ramachandran plot confirmed the conformation of the finest Helicase ATP binding domain (HABD) model anticipated by I-TASSER tool. The expected model's conformation was derived by means of examining the phi () and psi () torsion angles with the PROCHECK web service. PROCHECK's Ramachandran plot designates a high-grade model with over 90 percent residues in the maximum favored region [33].

Prediction of Secondary structure

The Helicase ATP binding domain (HABD) (187 amino acids) structure was demolished exhausting the phyre 2 server (http://www.sbg.bio.ic.ac.uk/phyre2). Phyre2 is a web-based tool for expecting & analyzing protein structure, function, & mutations [34]. A representative structural expectation will be returned between 30 minutes & 2 hours of submittal. The availability of beta strands, -helix, and disordered in the HABD region is substantial [20, 21].

Analysis of the HABD protein sequence and its content

A Zika virus helicase ATP binding domain amino acid sequence (1-187) was submitted to ExPASy Server via the link (https://web.expasy.org/protparam/), and data

was produced [35,36]. The whole amino acid content of the protein (in CSV format) is entered into an excel spreadsheet, and a graph is drawn to indicate the atomic design, which means the number of atoms in this protein (HABD). Similarly, we conducted extensive research to find positively and negatively charged amino acids. The sequence of helicase ATP binding protein was analyzed using the accessible link (NsitePred webserver. or http://biomine.cs.vcu.edu/servers/biomine.php?id=202 30714153327) for ligand prediction and critical probability with the score. The server is intended for predicting sequence-based binding residues for ATP, ADP, AMP, GDP, and GTP [37].

The kinase-specific phosphorylation site prediction

Phosphorylation is the best important posttranslational modification in eukaryotes, & it shows an essential role in various cellular functions. A computer prediction method is gaining popularity as a basic complementary strategy in researching protein phosphorylation sites. Prediction tools are classified into two types: Tools for kinase and non-kinase analysis (http://sysbio.unl.edu/PhosphoSVM). A kinase-specific expectation tool takes a protein sequence & the kinase type as input & outputs some measure of the probability that each S/T/Y residue in the sequence is phosphorylated via the chosen kinase [38- 41].

GPS 6.0 predicts protein/peptide phosphorylation

Protein phosphorylation, performed via protein kinases (PKs), is one of the essential PTMs and governs practically all biological activities. We used the FastA format of the HABD protein sequence to submit to the tool (https://gps.biocuckoo.cn/online.php) for p-site prediction. Then, choose the kinase node(s). The predictor has 10 S/T groups, 1 Y group, and 2 Dual groups for dual-specificity kinases. Press the "Submit" button and wait a bit. The extrapolation results of kinase-specific phosphorylation sites on the substrate are available. Protein statistics and disorder tendencies are also provided [42-44].

Results

The Zika Virus Genome Organization and its HABD Critical Domain

The genome of ZIKV (*Zika virus*) is a single-stranded (ss) positive-sense RNA molecule that comprises approximately 3423 amino acids of a polyprotein precursor [45]. It is hewn into three structural proteins (C, prM, and E) & eight nonstructural proteins (NS1, NS2A, NS2B, NS3, S4A, NS4B, & NS5) (Figure 1). The role of each domain (structural and nonstructural protein) is indicated in the domain architect below (left

side). The NS3 domain (Helicase domain) and its binding sites were the focus of this manuscript. The Helicase ATP binding domain (HABD) organizations were discovered utilizing available bioinformatics tools/servers. We found the HABD with 187 amino acid stretches (Figure 1). The HABD belongs to the DEAH-Box RNA helicase family. ATP binding site length (28-34 amino acids), DEAD box helicase motif length (117-120 amino acids). Similarly, we found the epitope peptides in the helicase ATP binding domain, depicted in blue and emphasized with blue color boxes (Figure 1).



Figure 1: Depicts the architecture of the Zika virus's helicase ATP binding (HABD) domain organization. The imperative conserved domains are written inside boxes and colored differently.

HABD protein 3-D structural analysis

I-TASSER successfully got the projected model of the Helicase ATP binding domain (HABD) protein & its 3-D coordinate file in PDB format. The server returned an anticipated secondary structure using a confidence score (ranging from 0 to 9), indicated solvent approachability, five anticipated facilities by way of Cscore, top ten PDB templates within alignment, top ten PDB structural, functional analogs protein, & binding site residues. Model HABD (Figure 2 A) was chosen as the finest C-score and RMSD model. C-scores with higher values indicate a higher-quality model. The top

ten threading templates for request protein sequence HABD were recognized by way of the LOMETS metaserver (Table 1). The threading alignment is commonly expected by normalized Z-score. However, а standardized Z-score >1 value reflects a confident alignment. Still, in the case of minor alignment of the extensive request sequence, it does not give a momentous sign of modeling accuracy. Then, the percentage sequence identity in the threading aligned region (Iden1) & the whole chain (Iden2) was careful about excellent homology (Table 1). With a TM-score of 0.883, the structural alignment program TM-align found that 2xauA in the PDB collection was the best structural analog of the highest-scoring model of I-TASSER (Table 2).

Rank	PDB Hit	Iden1	Iden2	Cov.	Z-score
1	3kx2B	0.21	0.26	0.97	2.55
2	2jlqA	0.73	0.60	0.82	2.75
3	6fa5A	0.19	0.24	0.90	2.99
4	6fszM	0.15	0.15	0.98	0.85
5	5vha	0.18	0.25	0.99	1.34
6	6fa5A	0.18	0.24	0.90	3.16
7	2v6j	0.75	0.64	0.84	0.99
8	2jlqA	0.71	0.71	1.00	3.01
9	5yvvB	0.77	0.59	0.78	2.25
10	1yksA	0.59	0.45	0.76	2.11

 Table 1: Shows the top ten threading alignment templates utilized by I-TASSER.

The sequence identity percentage of the templates in the threading aligned region with the query sequence is called Ident1. The percentage sequence identity of all the template chains with the query sequence is called Ident2. The threading alignment coverage is calculated by dividing the total number of aligned residues by the query protein's length. The threading alignments' normalized Z-score is known as the Z-score. A good alignment is indicated by an alignment with a Normalized Z-score >1 and vice versa.

Rank	PDB Hit	TM-score	RMSD ^a	IDEN ^a	Cov.	
1	2xauA	0.883	2.10	0.172	0.973	
2	6fa5A	0.882	2.13	0.172	0.973	
3	5xjcY	0.882	2.11	0.167	0.973	
4	5wsge	0.880	2.13	0.167	0.973	
5	6bk8P	0.874	2.25	0.168	0.968	
6	5xdrA	0.872	2.30	0.183	0.973	
7	5vhaA	0.866	2.29	0.168	0.984	
8	5aorA	0.865	2.55	0.152	0.984	
9	5n8rA	0.862	2.45	0.167	0.979	
10	6eudA	0.842	2.66	0.158	0.963	

Table 2: The PDB's top ten structural analogs were discovered by TM-alignment.

A scale for comparing two structures' structural similarity that was recently proposed is called the TMscore. The RMSD between residues that TM-align has structurally aligned is known as RMSDa. In the structurally aligned region, IDENa stands for the percentage sequence identity. Cov, the number of structurally aligned residues divided by the query protein's length, indicates the alignment's coverage by TM-align.

HABD protein predicted model evaluation

PROCHECK servers provided Ramachandran plots of the finest anticipated model, demonstrating the model's dependability. The PROCHECK Ramachandran plot revealed 95.1% residues in the most favored regions & 3.7% in further authorized areas, for a total of 98.8% residues in allowed regions (Figure 2 B).

HABD protein secondary structure prediction

Techniques for predicting secondary structures were evaluated. The Helicase ATP Binding Domain (HABD) has strand, -helix, and disordered secondary systems, according to the overall secondary structure investigation (Figure 2C). Elemental data, for example, secondary or tertiary structure predictions, can be used to characterize an underlying theory. Data on folds collected by biophysical techniques (e.g., circular dichroism) or functional data (existence of a conserved motif/domain) can also inform an initial strategy.

Analysis of HABD protein sequences and composition

A 187-amino acid protein sequence, including the Zika virus helicase ATP binding domain. The Protparam server was used to generate the amino acid sequence. We discovered the helicase ATP binding protein's atomic composition and several atoms. There are five atoms in total: nitrogen, sulfur, hydrogen, carbon, and oxygen. The graphical representation shows 1476 hydrogen atoms, 926 carbon atoms, 280 oxygen atoms, 256 nitrogen atoms, and ten sulfur atoms throughout the protein (Figure 3A). Finally, there are 2948 atoms in total. The hydrogen atom is essential in the composition of the helicase ATP binding domain protein, whereas sulfur plays a minor influence in its design. Similarly, we estimated the negatively and positively charged residues of this protein. We discovered that negatively charged amino acids (Asp + Glu) have a total of 26 and positively charged amino acids (Arg + Lys) have a capacity of 21 (Figure 3B). The molecular weight (20996.05) and theoretical Pi (5.46) were discovered. The entire amino acid content and several residues/parentages across the HABD protein are depicted in Figure 3C. Ala: 9.1%, Arg: 8.6%, Asn: 2.1%, Asp: 4.3%, Cys: 1.1%, Gln: 1.1%, Glu: 9.6%, Gly: 4.3%, His: 2.1%, Ile: 4.3%, Leo: 7.0%, Lys: 2.7%, Met: 4.3%, Phe: 3.2%, Pro:7.5%, Ser: 5.3%, Thr: 10.2%, Trp:

1.1%, Tyr: 2.7%, Val: 8. We also calculated the number of amino acids which are present in the HABD protein, such as A (17), R (16), N (4), D (8), C (2), Q (2), E (18), G (8), H (4), I (10), L (13), K (5), M (8), F (6), P (14), S (10), T (19), W (2), Y (5) and V(16) (Figure 3C). The graph displays the residue number and the query sequence, respectively, followed by the results of predictions for five ligand types: ATP, ADP, AMP, GDP, & GTP. Several predicted binding residues for HABD protein are ATP: 3, ADP: 8, AMP: 1, GTP: 0 & GDP: 6 (Figure 3D). Predictions for each ligand type include annotation of predicted binding residues & prediction scores, which estimate the probability of binding to a given ligand.



Figure 2: (A) (i) I-TASSER predicted the three-dimensional (3-D) structure of the Helicase ATP binding domain (HABD) protein. (ii) 2xauA template (iii) request protein (cyan) placed on structural analog (light blue) alignment 2xauA in the PDB library **(B)** PROCHECK Ramachandran plot of HABD confirmation of top score model of I-TASSER **(C)** Secondary structure prediction.

Identifying the phosphorylation location sites

In addition, we discovered a phosphorylation site in the HABD protein sequence. A protein sequence & kinase

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Similarly, with threonine, we created a sky-blue color graph with sequence, position, & scores. We found that threonine was present at 6, 22, 33, 47, 51, 57, 77, 78, 83, 87, 97, 99, 122, 135, 148, 150, 154, 168, 185, & locations with scores 0.924, 0.57, 0.353, 0.301, 0.681, 0.424, 0.456, 0.109, 0.108, 0.325, 0.122, 0.144, 0.521, 0.317, 0.324, 0.719, 0.378, 0.223, & 0.139 respectively (Figure 4D). We continuously show a green color plotted graph for tyrosine sequence, location, & scores. The results show that tyrosine is present at 75, 111, 114, 132, & 187, corresponding scores of 0.311, 0.465, 0.56, 0.533, and 0.273 (Figure 4E). Finally, we presented serine, threonine, and tyrosine p-sites and their sequence, position, & score.



Figure 3: The diagram of amino acid sequences Helicase ATP binding domain data was received after submitting it to the protparam server. **(A)** The diagram depicts the atomic structure of the protein. **(B)** This graph describes positively and negatively charged amino acid residues. **(C)** The overall amino acid composition of the protein and the number of residues/parentages. **(D)** The graph depicts the prediction of ligand type and binding probability.

Figure 4: (A) Different colors represent essential amino acids for phosphorylation. Red (serine), sky blue (threonine), and green (tyrosine) colors (B) The graph shows phosphorylation site predictions (serine, threonine, and tyrosine). (C) The red graph depicts the serine amino acid sequence, position, and score. (D) The graph in sky blue color depicts the threonine amino acid with sequence, position, and score. (E) The green graph depicts the tyrosine amino acid, sequence, position, and score.

GPS 6.0 is a group-based prediction system for envisaging PK-specific phosphorylation sites (p-sites) Protein phosphorylation is one of the most researched posttranslational modifications (PTMs) that affect biological processes [49]. We used ten-fold crossvalidation to test the concert of GPS 6.0 & each feature on pS/pT & pY sites for general p-site prediction. The sequences of the input can be submitted in FASTA format.



Figure 5: Protein/peptide phosphorylation prediction using GPS 6.0 **(A)** Expectation of kinase-specific phosphorylation site **(B)** Information on phosphorylation sites **(C)** Distribution of kinase family and p-sites **(D)** Annotations of the prediction results.

The expectation result provides possible p-sites by way of 11 different pieces of information, such as ID (the name/id of the protein sequence that you entered to forecast), Position (the location of the predicted phosphorylation site), (The residue that is expected to be phosphorylated), Kinase (the regulatory Kinase that is thought to phosphorylate the site), Score (The value calculated by the GPS 6.0 algorithm to assess phosphorylation potential. The greater the number, the more potentially phosphorylated the residue is), Cutoff (The cutoff value below the threshold. Different thresholds result in varying accuracy, sensitivity, and specificity) (Figure 5A). Surface accessibility and secondary structures, including -helix, -strand, & coil anticipated, could also be shown by NetSurfP [50] in the comprehensive mode, and they were established under the schematic figure (Figure 5B). We also ran basic statistics on the distribution of chosen kinase families, the number of disordered regions, & the secondary structures of anticipated p-sites (Figure 5C). All put in proteins' prediction results could be taken in one of four file formats: .txt, .csv, .tsv, & .xlsx. The 'Export' button assists in obtaining the annotation images.png file. A help page by way of an example input & output is also given (Figure 5D).

Discussion

After being isolated from Uganda for the first time in 1947, ZIKV eventually spread to other nations and regions. ZIKV has experienced several mutations and collected a variety of alterations in both structural and non-structural proteins, much like other RNA viruses [51]. Conversely, ZIKV strains that are circulating in Asian nations have the 139S residue in the preM protein, which has been linked to ZIKV-related microcephaly on occasion [52]. One element of the ZIKV replication complex is NS3. Strong contacts between the N-terminal of NS3 and the hydrophilic portion of the integral membrane protein NS2B bind it in the extracellular space. The helicase domain of the NS3 C-terminal domain is known to have NTPase and helicase activity.

The N-terminal and its cofactor NS2B combine to generate a protease that cleaves the flavivirus polyprotein. ZIKV NS3 helicase, part of the helicase superfamily 2 (SF2), is the ZIKV helicase/NTPase domain. It functions with NS5 polymerase to unwind organized template sections of RNA during the de novo viral RNA synthesis process. In addition, RNA binding stimulates the nucleoside triphosphatase activity of NS3 helicase, which supplies the chemical energy required to unwind RNA intermediates during viral RNA replication [53, 54]. They were observed when the GDP and GTP models accurately predicted the residues that bind GTP and GDP, respectively. This implies that among the predictions produced by NsitePred, structural similarity between the ligands-which influences similarity in their binding is also noted. However, the recall is significantly lower when a particular model is used to predict binding residues for less comparable nucleotides, such as when NsitePred_ATP predicts the GDP, GTP, and AMP-

binding chains. This suggests that variations in nucleotide structures lead to variations in the related prediction models, which in turn drives the creation of consensus-based predictors [55]. In this study, GPS 6.0 provides PK-specific p-site predictions for 44 046 PKs across 185 species that are believable. With this assistance, more users could forecast p-sites and upstream kinases on substrates in the relevant study fields. To interpret the p-sites and substrates, we additionally integrated a lot of visible annotations. The prediction results would be given more thoroughly with the p-site and substrate information included. A userfriendly web server with highly adjustable ssKSRs prediction, selectable kinase, and various modules, GPS 6.0, was built [56]. The current study uses GPS 6.0 for amino acid composition, phosphorylation sites, genome analysis, and structure prediction.

In tropical & subtropical areas, the Aedes genus mosquito, mainly *Aedes aegypti*, is the principal vector for transmitting the Zika virus. The highest times for Aedes mosquito bites throughout the day are in the early morning & late afternoon, and early evening. The mosquito that spreads dengue, chikungunya, & yellow fever is the same. Using in-silico genome prediction, other domain organizations. We concentrate on the NS3 domain, the only domain with helicase unwinding and ATP binding capabilities. Additionally, we forecast helicase ATP-binding domain epitope-binding peptides. In parallel, using the Ramachandran plot, we performed structure modeling to validate. We have performed atom-composition studies of proteins, including positively and negatively charged amino acids, to gain insight into the in-depth study. We have also predicted the protein's phosphorylation sites (psites) and numerous p-sites. In the future, more results-interpreting annotations will be gathered from further open sources & tools. Additionally, we'll add more improvements to boost performance. For academic research, GPS 6.0 will be uninterruptedly sustained & amended.

Author Contributions

AS and NA, prepare the first draft, wrote the MS.

Conflict of Interest

The authors declare that there is no conflict of interest.

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